



**Proposal for Task Force Consideration
at the ISSC 2019 Biennial Meeting**

1. a. Growing Area
 b. Harvesting/Handling/Distribution
 c. Administrative

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10. Proposal Subject	Laboratory Method for <i>Vibrio parahaemolyticus</i> and <i>Vibrio vulnificus</i> Enumeration and Detection Through MPN and Real-Time PCR
11. Specific NSSP Guide Reference	Section IV Guidance Documents Chapter II Growing Areas .14 Approved NSSP Laboratory Tests

12. Text of Proposal/ Requested Action	5. Approved Methods for <i>Vibrio</i> Enumeration			
		Vibrio Type:	Application: PHP Sample Type: Shucked	Application: Reopening
	EIA ¹	<i>Vibrio vulnificus (V.v.)</i>	X	
	MPN ²	<i>Vibrio vulnificus (V.v.)</i>	X	
	SYBR Green 1 QPCR-MPN ⁵	<i>Vibrio vulnificus (V.v.)</i>	X	
	MPN ³	<i>Vibrio parahaemolyticus (V.p.)</i>	X	
	PCR ⁴	<i>Vibrio parahaemolyticus (V.p.)</i>	X	
	MPN-Real Time PCR ⁶	<i>tdh+ and trh+ Vibrio parahaemolyticus (V.p.)</i>	X	X
	MPN-Real Time PCR ⁷	<i>Vibrio parahaemolyticus (V.p.)</i>	X	X
	<u>MPN-Real Time PCR⁹</u>	<u><i>Vibrio parahaemolyticus (V.p.) and Vibrio vulnificus (V.v.)</i></u>	<u>X</u>	<u>X</u>
Direct Plating Method ⁸	<i>Vibrio parahaemolyticus (V.p.)</i>	<u>X</u>	X	
<p>Footnotes: ¹ EIA procedure of Tamplin, et al, as described in Chapter 9 of the FDA Bacteriological Analytical Manual, 7th Edition, 1992.</p>				

	<p>² MPN method in Chapter 9 of the FDA Bacteriological Analytical Manual, 7th Edition, May 2004 revision, followed by confirmation using biochemical analyses or by the DNA -alkaline phosphatase gene probe for <i>vvhA</i> as described by Wright et al., or a method that a State can demonstrate is equivalent.</p> <p>³ MPN method in Chapter 9 of the FDA Bacteriological Analytical Manual, 7th Edition, May 2004 revision, followed by confirmation using biochemical analyses or the DNA-alkaline phosphatase gene probe for <i>tlh</i> as described by McCarthy et al., or a method that a State can demonstrate is equivalent.</p> <p>⁴ MPN method in Chapter 9 of the FDA Bacteriological Analytical Manual, 7th Edition, May 2004 revision, and as described in the “Direct Plating Procedure for the Enumeration of Total and Pathogenic <i>Vibrio parahaemolyticus</i> in Oyster Meats” developed by FDA, Gulf Coast Seafood Laboratory, or a method that a State can demonstrate is equivalent.</p> <p>⁵ <i>Vibrio vulnificus</i>, ISSC Summary of Actions 2009. Proposal 09-113, Page 123.</p> <p>⁶MPN-Real Time PCR Method for the <i>tdh</i> and <i>trh</i> Genes for Total <i>V. parahaemolyticus</i> as described in Kinsey et al., 2015. ISSC 2015 Summary of Actions Proposal 15-111, Page 397.</p> <p>⁷MPN-Real Time PCR Method for the <i>tlh</i> gene for total <i>V. parahaemolyticus</i> as described in Kinsey et al., 2015. ISSC 2015 Summary of Actions Proposal 15-113, Page 418</p> <p>⁸Direct Plating Procedure in Chapter 9 of the FDA Bacteriological Analytical Manual, 7th Edition, May 2004 revision, and as described in the ‘Direct Plating Procedure for the Enumeration of Total and Pathogenic <i>Vibrio parahaemolyticus</i> in Oyster Meats’ developed by FDA, Gulf Coast Seafood Laboratory.</p> <p><u>⁹MPN-Real Time PCR Method for <i>Vibrio parahaemolyticus</i> and <i>Vibrio vulnificus</i>. Washington State Department of Health, Food and Shellfish Bacteriology Laboratory.</u></p>
<p>13. Public Health Significance</p>	<p>The purpose of this method is to provide laboratories supporting the NSSP the ability to rapidly quantify <i>Vibrio parahaemolyticus</i> (<i>Vp</i>) and <i>Vibrio vulnificus</i> (<i>Vv</i>) from oysters using a high throughput real-time PCR assay. Rapid and early detection of these pathogens, complying with the required quantitative detection guidelines suggested by the ISSC, will help the shellfish industry market oysters for consumption that are within regulatory limits for these pathogens.</p> <p>This method once approved would add a testing method of MPN Real-Time PCR for <i>Vibrio vulnificus</i> and it would be an alternative to the <i>Vibrio parahaemolyticus</i> MPN Real-Time PCR methods already approved in the 2017 Model Ordinance.</p>
<p>14. Cost Information</p>	<p>The cost for this method is approx. \$155 per sample. This estimate is based on recurring costs of consumables, reagents, and supplies needed for routine testing. It does not include indirect materials considered to be standard microbiology equipment such as analytical balance, PCR workstation, DNA purification system, refrigerator, pipettes, etc.</p>