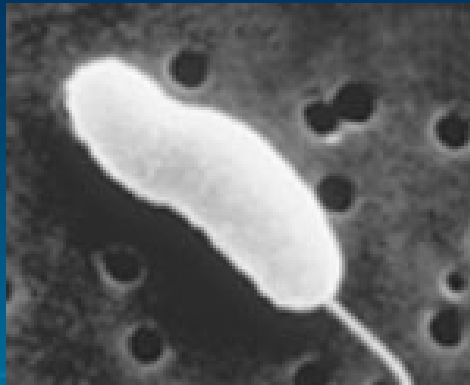


Rapid Detection Methods: An Industry Necessity, A Laboratory Challenge!

M. Lizzie Evans
August 18, 2010



OVERVIEW

- SHORT Introduction to *Vibrios*
- PCR lesson and options
- Laboratory implications of molecular methods; checklists and methods

Vibrio vulnificus: An Introduction

- CDC (1997-2004): *Vibrio* most frequently isolated genus in Gulf coast waters. (CDC, 2005)
- *Vibrio vulnificus*
 - Leading cause of mortality from seafood consumption (fatality rate > 50%)
 - Low infectious dose; most severe foodborne infections- but only in susceptible persons
 - 80% of infections between May and October

Motes et al, 1998; Hlady et al, 1993; Kelley et al, 1997

Wound infection vs. septicemia

- Wounds from exposure to shellfish or seawater
- Septicemia follows ingestion of raw oysters
- Post-harvest processing (PHP) of oysters mandated by FDA?
- Rapid detection is crucial for the consumer and the industry



Post-Harvest Processing of Oysters

- PHP- summer months
 - *For cooking? For shucking? For PHP?
- Demonstrate reduction of *V. vulnificus* by achieving 3.52 log reduction
- After validation of method, “vibrio reduced”. Verified monthly.

The Past and the Future for the Shellfish Industry

- PHP to reduce *V. vulnificus* in 25% of oyster shellstock from Gulf Coast (2003)
- PHP for *V. parahaemolyticus* (2005)
- California bans untreated oysters (2005)
- Monitor harvest waters for Vibrios (2007)
- Demonstrate 60% reduction in *V. vulnificus* disease by 2010 (2007)

The Past and the Future for the Shellfish Industry

- Rescinded Ban? well....
- Time/temperature controls
- What's are we doing?

What Is In Place Right Now?

- *Vibrio* infections have continued to occur in spite of the efforts of regulatory agencies, industry and the scientific community.
- 2007, national surveillance system for *V. vulnificus*. CDC collaborated with Alabama, Florida, Louisiana, Texas, and Mississippi to monitor cases of *V. vulnificus* infection in the Gulf Coast region; nationally notifiable.
- 2010, Integrated (Federal/State) Food Safety System: Reportable Food Registry

Detection Options

- Biochemicals... BAM
- DNA Probe
- PCR options, either approved or in the pipeline (hint: multiplex)...

Validating Methods: PHP for industry



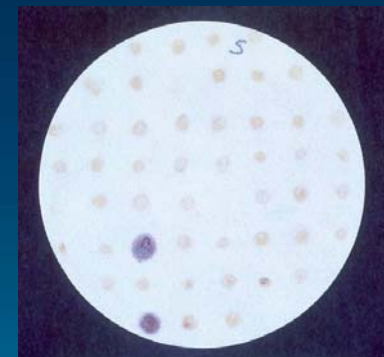
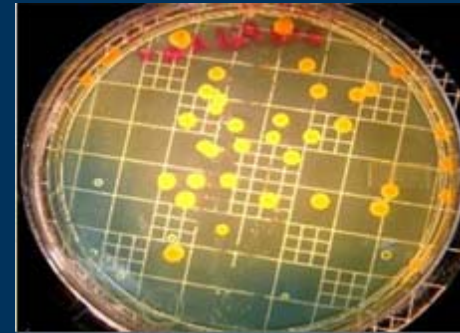
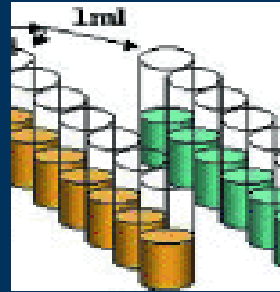
- FDA-MPN (FDA, 2004)
 1. Homogenized and inoculated into MPN tubes (24hrs)
 2. Streaked on selective media (24hrs)
 3. Picked and spotted onto nutritive media (24hrs)
 4. Lifted and pro-k (24 to 48hrs)
 5. Hybridized (<24 hrs).

- This process is as tedious as it sounds!!! So scientists at UF did something about it...

QPCR Application for MPN



1. Homogenize oysters and dilute
2. Replicates (n=3-10) of APW enrichment tubes for MPN (Day 1)
3. Streak to selective agar (Day 2)
4. Pick colonies (Day 3)
5. Identify colonies by DNA probe (Day 4)

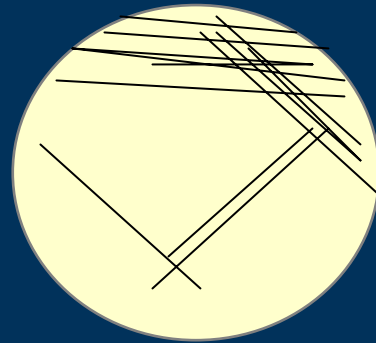


1. OR... PCR

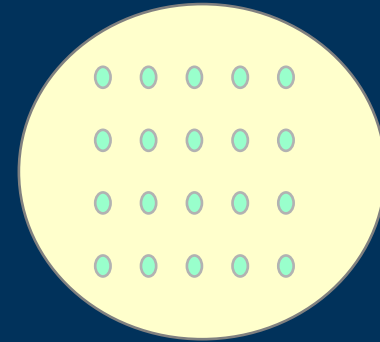
Colony Blot Hybridization



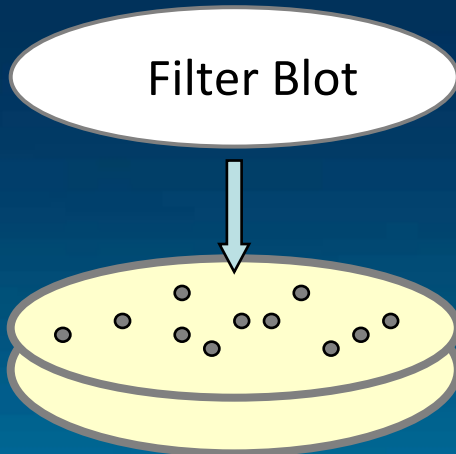
Culture



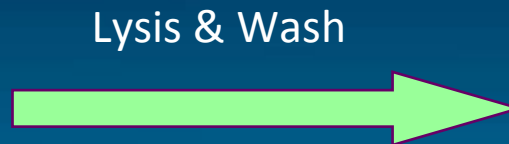
Streak to selective



Spot positive colonies



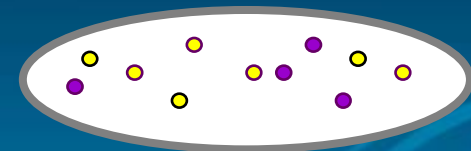
Filter Blot



Lysis & Wash

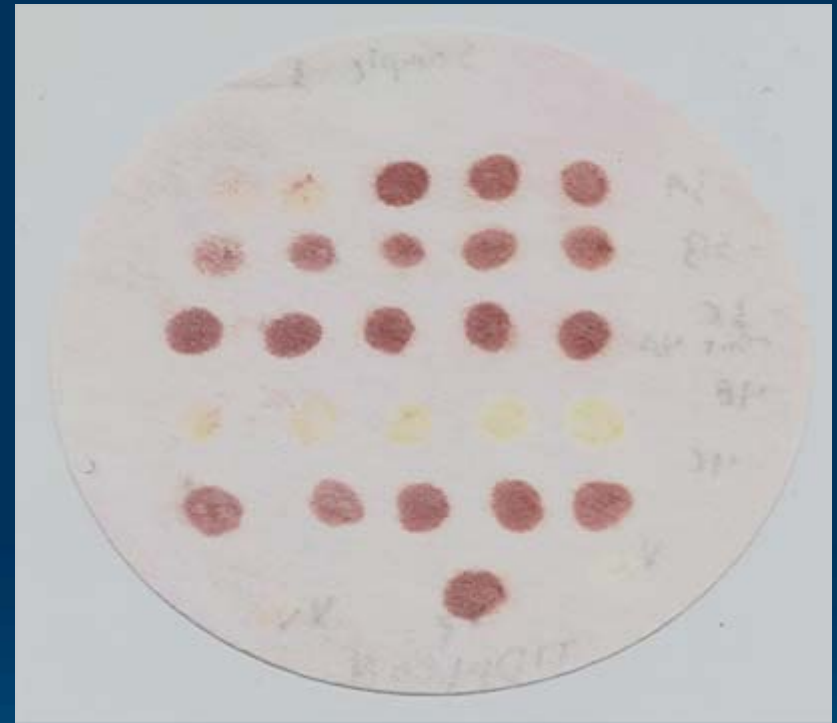
Hybridization

Development



Probed Filter with positive and negative colonies

- Purple (+)
- Yellow (-)
- VERY EXPENSIVE!
- Can be subjective...

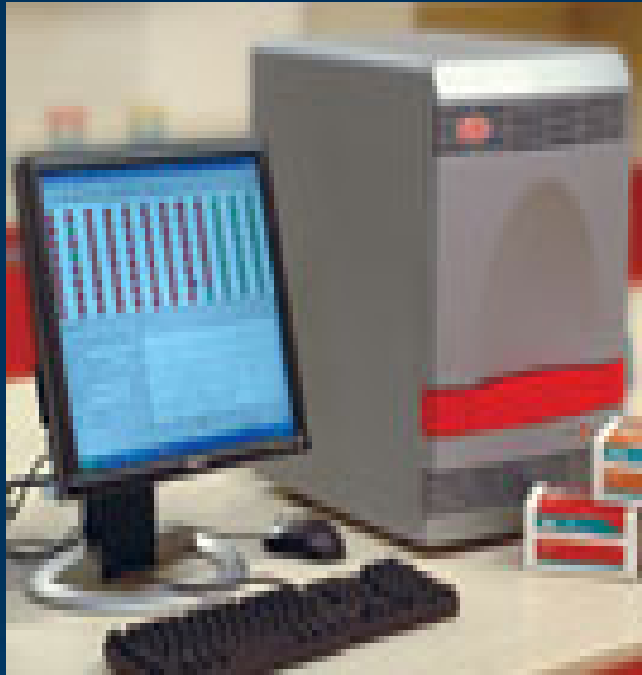


Yes we can.....PCR

- Mastermix: primers, taq, dNTP's (ACTG's), Sybr green dye 1, water and salt
- Standard curve to quantify with known concentrations of *V.v.*
- Result in CT value and Melt temp.
- WE HAVE A CHECKLIST COMING AND A METHOD IN NSSP!!!



...Confirm positive APW directly by Q-PCR



DuPont Qualicon Q7

Cost?



Cepheid Smartcycler

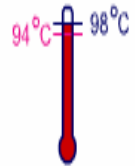
How PCR is done!

<http://www.sigmaaldrich.com/lifescience/molecularbiology/pcr/learning-center/sybr-green-animation.html>

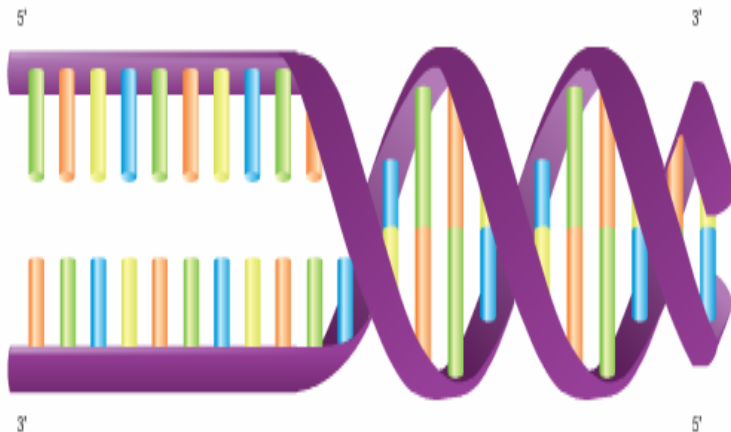


CRanimation.mp

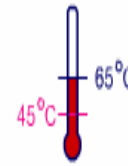
Denaturation -



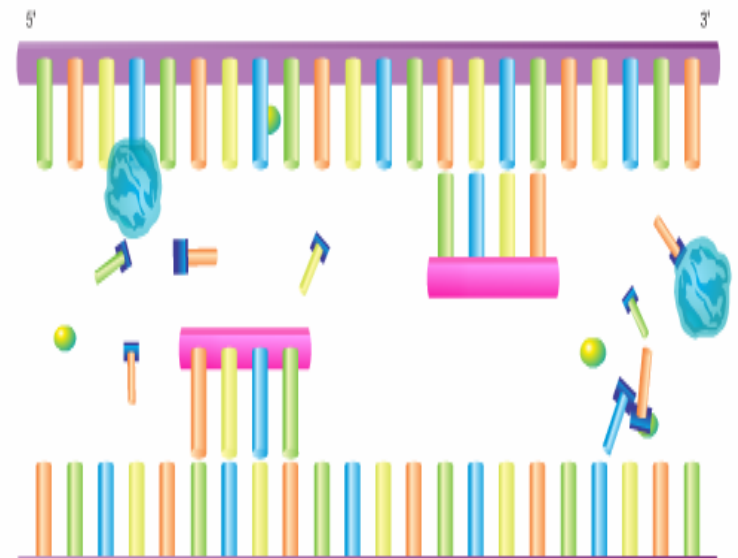
Double-stranded DNA is heated to 94°C - 98°C. During this period, the double-stranded DNA helix melts open into two single-stranded DNA templates.



Annealing -

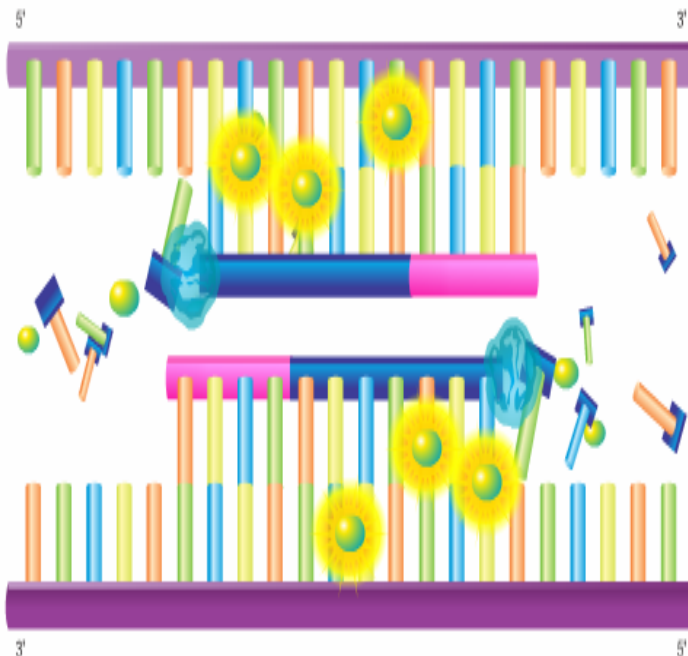
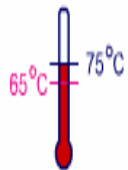


The reaction is cooled to 45°C - 65°C. Single-stranded primers anneal to the single-stranded DNA template. During this cycle, DNA polymerase attaches to the primed template and begins to incorporate complementary nucleotides (dATP, dCTP, dGTP, TTP). This process is very slow because the polymerase is inefficient at these lower temperatures.

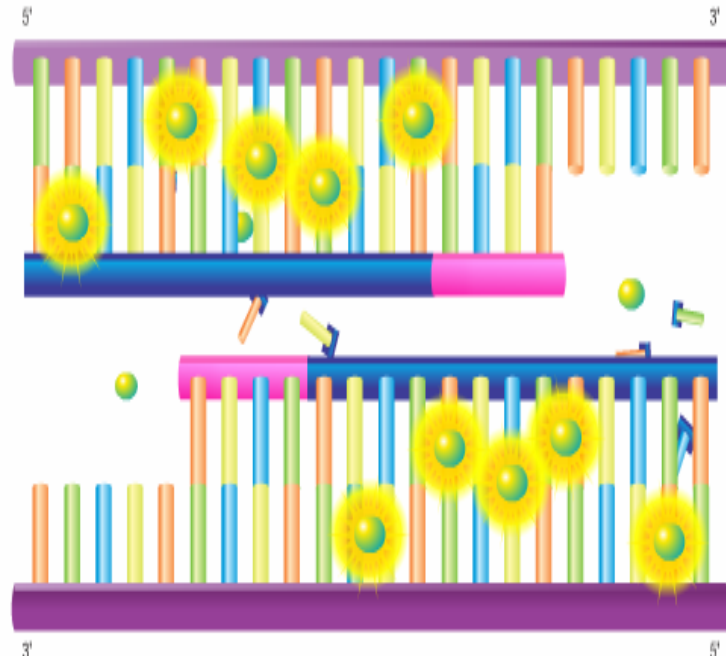


How PCR is done: part 2

Extension - SYBR Green I binds all newly synthesized double-stranded DNA complexes and fluoresces. The fluorescence accumulates as cycling of PCR continues and is measured at the end of each PCR cycle. The intensity of fluorescence generated by SYBR Green I above background level (C_T value) is measured and used to quantitate the amount of newly generated double-stranded DNA strands.



Replicated DNA - After repeating the denaturation, annealing, and extension cycles approximately 35-40 times, you are ready to begin analysis. The C_T values can be used to quantitate starting amounts of DNA or to establish a standard curve for gene expression studies or other comparative analysis.



Sybr method: Cepheid®

➤ Method utilized in
V. vulnificus SYBR
method

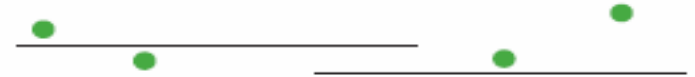
➤ ISSC interim
approval

SYBR® GREEN I DYE ASSAY CHEMISTRY

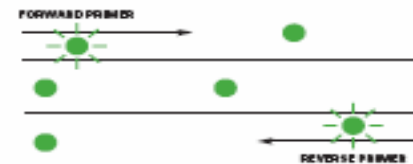
1. **Reaction setup:** The SYBR® Green I Dye fluoresces when bound to double-stranded DNA.



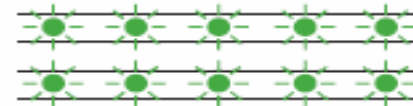
2. **Denaturation:** When the DNA is denatured, the SYBR® Green I Dye is released and the fluorescence is drastically reduced.



3. **Polymerization:** During extension, primers anneal and PCR product is generated.

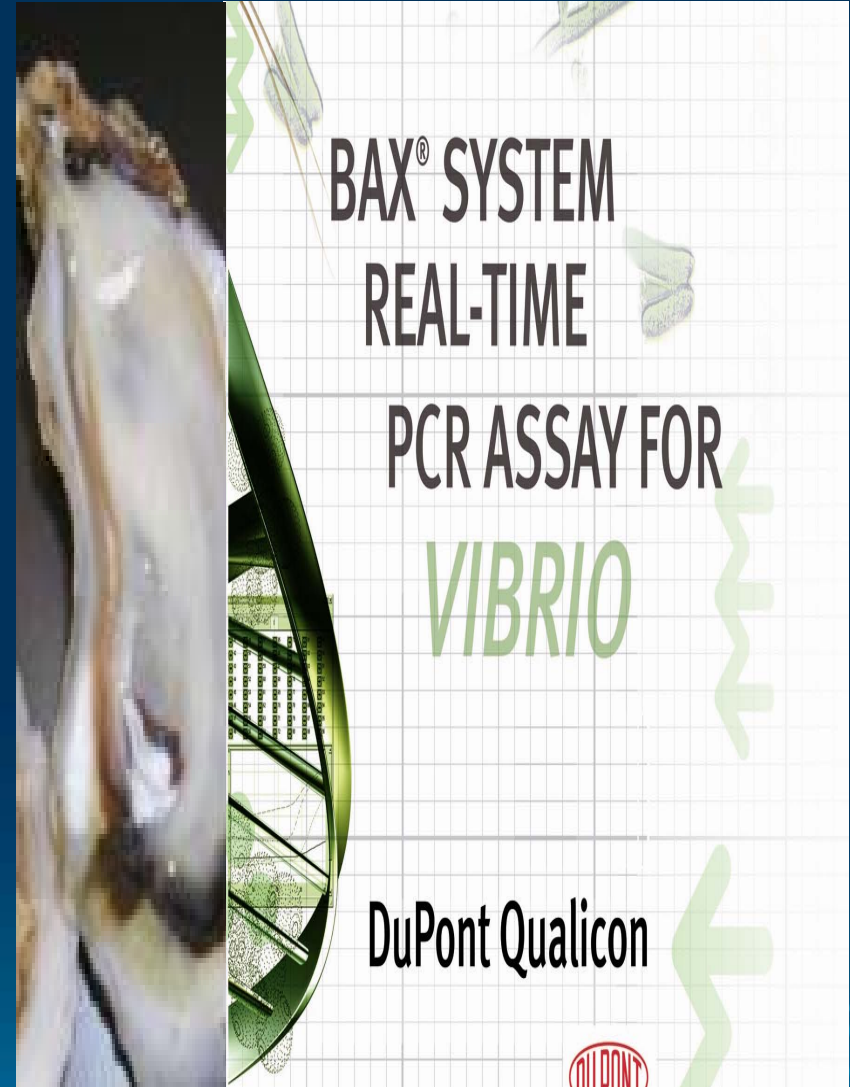


4. **Polymerization completed:** When polymerization is complete, SYBR® Green I Dye binds to the double-stranded product, resulting in a net increase in fluorescence detected by the 7900HT system.




DuPont method currently under consideration

- Frequently utilized, probes for quantitative PCR.
- Multiplex; *V.v.* *V.p.* *V.c.*
- TaqMan-based QPCR
- Simplified extraction
Internal control standard
- AOAC approved to 10^4 CFU/ml
- Protocol submitted to ISSC

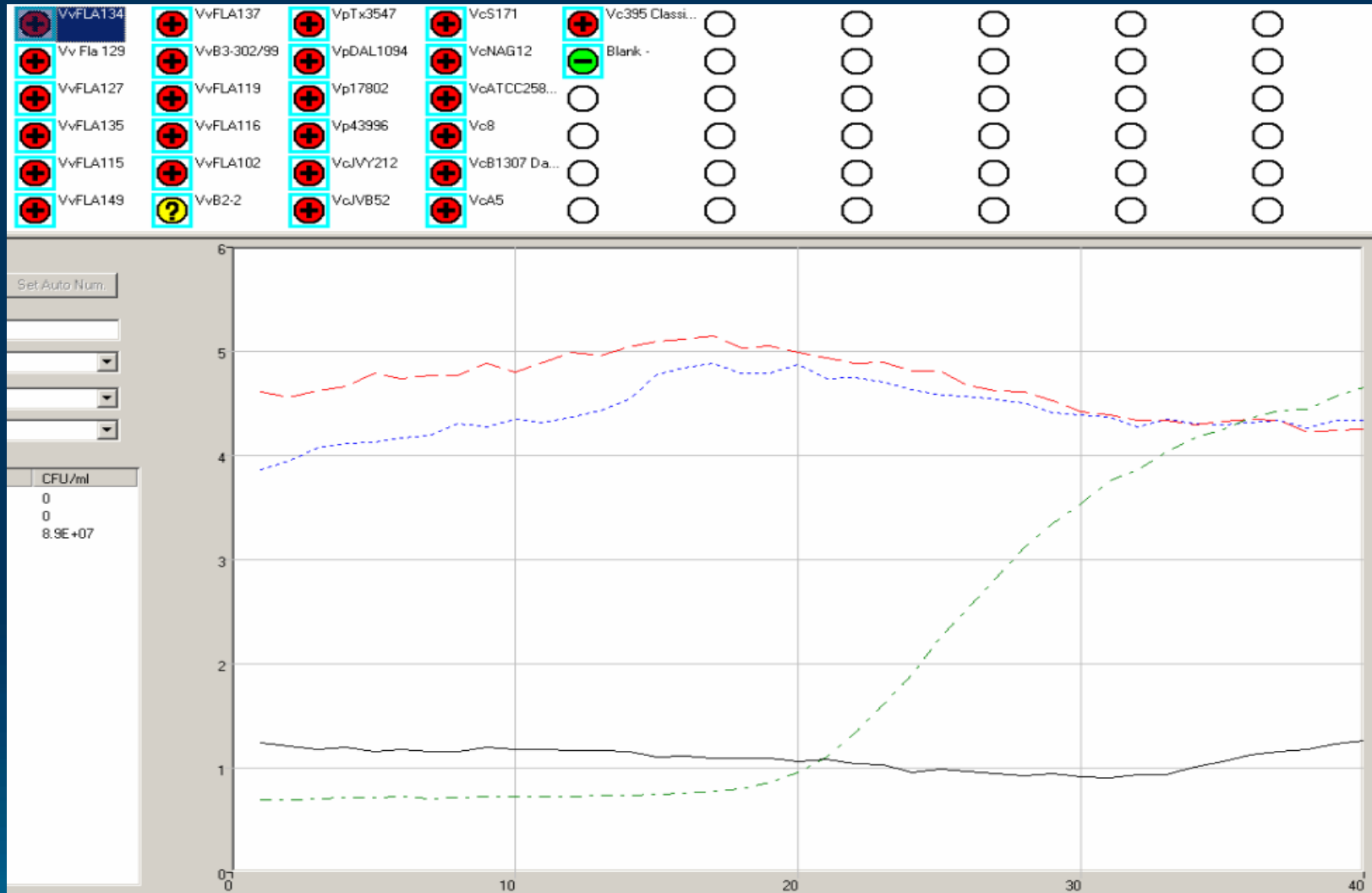


BAX[®] SYSTEM
REAL-TIME
PCR ASSAY FOR
VIBRIO

DuPont Qualicon



Vibrio Multiplex (Dupont Qualicon)



Checklist Information

➤ Critical Information:

➤ All procedures, with materials, methods, calibrations and directive regarding interpretations of data.

➤ Quality information is similar, points of interest for PCR

➤ Sybr method standard curve and interpretation...
DuPont method no interpretation.

➤ Commercial kit.

NSSP/BAM methods.... How to implement in your lab (AKA what the checklist says)

- Must have a PCR area that does not contain live cultures
- Must have cold storage available
- Either platform is costly!
- Cepheid: thermocycler, centrifuge, heat block etc
- DuPont: thermocycler and heatblocks included. No centrifugation...

Validating and verifying methods the easy way!!



- Quantitative PCR was evaluated, as it may increase both speed with which samples may be processed without sacrificing sensitivity.
- Homogenized and inoculated into MPN tubes (24hrs) or possible to run the sample without enrichment! Extract and run PCR (<24 hrs)

I Want to Validate a Method. What Do I Do Next?

- New guidance documents under review to standardize submission.
- Schedule an evaluation for the laboratory involved in the microbiological testing
- Call and ask!!

Validation Results

- Results of a side by side sample comparison support application of Q-PCR technology for validation oyster processing protocols
- Quantitative PCR was applied to most probable number (QPCR-MPN) for validation of PHP methods for reduction of *Vibrio vulnificus* in oysters.

So What???

- Industry needs an efficient method of detection, especially with trend of PHP mandates.
- Laboratories will be implementing molecular methods and training and guidance will be available through FDA and LEO's
- Checklists are currently in final revisions and will be submitted to ISSC in 2011 for acceptance.
- Molecular methods will allow industry an

References

1. CDC (2005). Center for Disease Control. Annual Summaries of Surveillance of Outbreaks of *Vibrio* infection, 1997 - 2004.
2. Gulig, P. A., K. L. Bourdage, and A. M. Starks. 2005. Molecular pathogenesis of *Vibrio vulnificus*. *J. Microbiol.* 43:118-131.
3. Wright, A.C., L.M. Simpson, and J.D. Oliver, Role of iron in the pathogenesis of *Vibrio vulnificus* infections. *Infect Immun*, 1981. 34(2): p. 503-7.
4. Garthright, R. J. Blodgett, and S. J. Chirtel. 1998. Influence of water temperature and salinity on *Vibrio vulnificus* in Northern Gulf and Atlantic Coast oysters (*Crassostrea virginica*). *Appl. Environ. Microbiol.* 64:1459–1465
5. Hlady, W.G., R.C. Mullen, and R.S. Hopkin, *Vibrio vulnificus* from raw oysters. Leading cause of reported deaths from foodborne illness in Florida. *J Fla Med Assoc*, 1993. 80(8): p. 536-8.
6. Kelley, J.I., et al., Effects of Temperature, Salinity, and Substrate on the Colonization of Surfaces In Situ by Aquatic *Bdellovibrios*. *Appl Environ Microbiol*, 1997. 63(1): p. 84-90.
7. Interstate Shellfish Sanitation Conference. 2003. Issue relating to a *Vibrio vulnificus* risk management plan for oysters. ISSC, Columbia, S.C.
8. Tacket CO, Brenner F, Blake PA. Clinical features and an epidemiological study of *Vibrio vulnificus* infections. *J Infect Dis* 1984;149:558-61.
9. CDC. *Vibrio vulnificus* infections associated with raw oyster consumption -- Florida, 1981-1992. *MMWR* 1993;42:405-7.
10. FDA, Food and Drug Administration. FDA warning on raw oysters. Food and Drug Administration. *AIDS Treat News*, 1995(no 229): p. 6.
11. FDA. (1997). National Shellfish Sanitation Program (NSSP), Guide for the Control of Molluscan Shellfish, Model Ordinance.
12. FDA. (2000). National Shellfish Sanitation Program (NSSP), Guide for the Control of Molluscan Shellfish, Model Ordinance revision. <http://www.cfsan.fda.gov/~ear/nsspotoc.html>

References continued

13. G.O.I.C. (2001) Gulf Oyster Industry Leads the Way Toward Providing Less Risky Oysters for Certain At Risk Consumers Through Post-Harvest Treatment. PHT Products Volume,
14. Harwood, V. J., J. P. Gandhi, and A. C. Wright. 2004. Methods for isolation and confirmation of *Vibrio vulnificus* from oysters and environmental sources: a review. *J Microbiol Methods* 59:301-311.
15. (FDA), U.S.F.a.D.A., Cooperative Agreement to Support the Shellfish and Seafood Safety Assistance Project D.o.H.a.H. Services, Editor. 2006.
16. M. J. Espy,* J. R. Uhl, L. M. Sloan, S. P. Buckwalter, M. F. Jones, E. A. Vetter, J. D. C. Yao, N. L. Wengenack, J. E. Rosenblatt., Real-Time PCR in Clinical Microbiology: Applications for Routine Laboratory Testing. *CLINICAL MICROBIOLOGY REVIEWS*, Jan. 2006, p. 165–256 Vol. 19, No. 1
17. FDA, Bacteriological Analytical Manual Online. Chapter 9, 2004(In A. D. AKaysner CA ed.).
18. Blackstone, G.M., et al., Detection of pathogenic *Vibrio parahaemolyticus* in oyster enrichments by real time PCR. *J Microbiol Methods*, 2003. 53(2): p. 149-55.
19. Wright, A.C., et al., Evaluation of Post-Harvest Processed Oysters using PCR-based Most Probable Number for *Vibrio vulnificus*. *Appl. Environ. Microbiol.*, 2007: p. AEM.01118-07.
20. Campbell, M.S. and A.C. Wright, Real-time PCR analysis of *Vibrio vulnificus* from oysters. *Appl Environ Microbiol*, 2003. 69(12): p. 7137-44.
21. Blackstone, G.M., et al., Use of a real time PCR assay for detection of the *ctxA* gene of *Vibrio cholerae* in an environmental survey of Mobile Bay. *J Microbiol Methods*, 2007. 68(2): p. 254-9.
22. Nordstrom, J.L., et al., Development of a Multiplex Real-Time PCR Assay with an Internal Amplification Control for the Detection of Total and Pathogenic *Vibrio parahaemolyticus* Bacteria in Oysters. *Appl. Environ. Microbiol.*, 2007. 73(18): p. 5840-5847.
23. CDC. MMWR. Morbidity, Mortality, and Mollusks: *Vibrio vulnificus* Infections Associated with Raw Oyster Consumption - California, 1991-2001.

QUESTIONS????



Lizzie Evans: